ELSEVIER

Contents lists available at ScienceDirect

## **Bioorganic & Medicinal Chemistry Letters**

journal homepage: www.elsevier.com/locate/bmcl



# 3-(3-Aryloxyaryl)imidazo[1,2-a]pyridine sulfones as liver X receptor agonists

Robert R. Singhaus <sup>a,\*</sup>, Ronald C. Bernotas <sup>a</sup>, Robert Steffan <sup>a</sup>, Edward Matelan <sup>a</sup>, Elaine Quinet <sup>b</sup>, Ponnal Nambi <sup>b</sup>, Irene Feingold <sup>c</sup>, Christine Huselton <sup>c</sup>, Anna Wilhelmsson <sup>d</sup>, Annika Goos-Nilsson <sup>d</sup>, Jay Wrobel <sup>a</sup>

- <sup>a</sup> Chemical Sciences, Wyeth Pharmaceuticals, 500 Arcola Rd., Collegeville, PA 19426, USA
- <sup>b</sup> Cardiovascular and Metabolic Diseases, Wyeth Pharmaceuticals, 500 Arcola Rd., Collegeville, PA 19426, USA
- <sup>c</sup> Drug Safety and Metabolism, Wyeth Pharmaceuticals, 500 Arcola Rd., Collegeville, PA 19426, USA
- d Karo Bio AB, Novum S-141, 57 Huddinge, Sweden

#### ARTICLE INFO

Article history:
Received 15 October 2009
Revised 18 November 2009
Accepted 19 November 2009
Available online 23 November 2009

Keywords: Imidazo[1,2-a]pyridine Liver X receptor LXR Atherosclerosis Sulfone ABCA1 Cholesterol Triglycerides

#### ABSTRACT

Replacement of a quinoline with an imidazo[1,2-a]pyridine in a series of liver X receptor (LXR) agonists incorporating a [3-(sulfonyl)aryloxyphenyl] side chain provided high affinity LXR ligands 7. In functional assays of LXR activity, good agonist potency and efficacy were found for several analogs.

© 2009 Elsevier Ltd. All rights reserved.

Liver X receptors (LXRs) are members of the nuclear receptor superfamily. 1 LXRs act as ligand activated transcription factors which increase the expression of several genes when turned on by agonists including oxysterols, the endogenous agonists. The activation requires the formation of an LXR-RXR (retinoid X receptor) heterodimer. Among the key genes activated by LXR-RXR heterodimers are those corresponding to the ATP-binding cassette proteins (ABCs), a group of lipid transporters partly responsible for controlling lipid homeostasis.2 Increasing the expression of ABCs and in particular ABCA1, an important transporter in macrophages and other cells, may increase reverse cholesterol transport (RCT). Enhancing RCT could have the effect of slowing the development of atherosclerosis. However activation of undesired genes, especially sterol regulatory element-binding protein-1c (SREBP-1c) and fatty acid synthetase (FAS), also under the control of LXRs, may lead to increases in triglycerides (TGs) and to steatosis in the liver.<sup>3</sup> To minimize the potential for undesired effects, agonism of the LXR<sub>\beta</sub> subtype (present in macrophages and independently able to upregulate ABCA1) rather than of the  $\alpha$ -subtype (more prevalent

As an alternative or addition to statin therapy, several companies have targeted LXR agonists for dislipidemia. These include T0901317 (1)<sup>5</sup> from Tularik and GW3965 (2)<sup>6</sup> from GlaxoSmithK-line (Fig. 1). These compounds are high affinity LXR ligands with potent LXR agonism. An indazole-based LXR agonist, WAY-252623 (3),<sup>7</sup> has been shown to upregulate ABCA1 in whole blood of cynomolgus monkeys<sup>8</sup> and like wise in humans in a phase I clinical trial (Fig. 2).<sup>9</sup> A quinoline-based series also developed by Wyeth is exemplified by WAY-254011 (4).<sup>10</sup>

Figure 1. LXR Agonists from Tularik and GlaxoSmithKline.

in liver than LXR $\beta$  and the site of TG synthesis) has been the focus for several groups.<sup>4</sup>

<sup>\*</sup> Corresponding author. E-mail address: Singhar@wyeth.com (R.R. Singhaus).

Figure 2. LXR Agonists from Wyeth.

The moderate peroxisome proliferator-activated receptor (PPAR) agonism of quinoline  $\bf 4$ , which activated all three subtypes of the receptor,  $^{11}$  led us to target quinolines with a biarylether substituent and to identify non-carboxylic acid hydrogen bond acceptors,  $^{12}$  leading to 4-[(3-sulfone-phenoxy)phenyl]quinolines  $\bf 5$ . These compounds were essentially devoid of PPAR agonism. To further develop the SAR, we explored replacements for the quinoline such as benzimidazole which maintained a key sp $^2$  nitrogen and trifluoromethyl ( $\bf Z = CF_3$ ) on the core heterocycle ( $\bf 6$ ). As an extension of this approach, we describe here the synthesis and structure-activity relationships of a series of imidazo[1,2-a]pyridines  $\bf 7$ .

The first approach to targets **7** involved ether bond formation between a 3-(3-hydroxyphenyl)imidazo[1,2-a]pyridine core **11** and an arylhalide **12** (Scheme 1). The phenols were typically prepared by reaction of 2-aminopyridines **8** with  $\alpha$ -haloketones **9** to give imidazo[1,2-a]pyridines **10**<sup>15</sup> in which substituents Y and Z were varied by appropriate choice of starting materials. <sup>16</sup> Since

preparation of  $PhCH_2C(O)CH_2Cl$  (9, Y = benzyl) was problematic, imidazo[1,2-a]pyridine **10b** (Y =  $CH_2Ph$ , Z =  $CF_3$ ) was made from 10a by standard Suzuki coupling with phenylboronic acid (Scheme 2).<sup>17</sup> Direct arylation of compounds **10** with 3-iodophenol using Pd(OAc)<sub>2</sub><sup>18</sup> or more conveniently with Pd(OH)<sub>2</sub> on carbon<sup>19</sup> gave phenols 11 in good yield. Introduction of the sulfone or sulfonamide side chains by reaction with arylhalide 12<sup>20</sup> was accomplished either through copper-mediated coupling with aryliodides and arylbromides<sup>21</sup> or by aromatic substitution for arylfluorides and arylchlorides.<sup>22</sup> Incorporation of amines and azacycles on a methylene group at the 2-position required bromination of 2methyl analog 7a to give 7j followed by displacement reactions to give 7k-70 (Scheme 3). When sulfonamides were prepared, para-methoxybenzyl protection<sup>23</sup> was employed to provide 7s and 7t which were deprotected with trifluoroacetic acid to give primary and secondary sulfonamides 7u and 7v, respectively.

Alternative approaches to **7** were used to explore the structure-activity relationships, especially for various linkers between the aromatic rings of the side chain other than oxygen (Scheme 4). For example, synthesis of compounds with biaryl side chains in which the aromatic rings were connected by other linkers (L) including methylene (**7x**) and oxymethylene (**7y**) was accomplished by making arylbromides such as **14**<sup>24</sup> and **17** for use in the direct arylation. Ultimately, coupling a preformed biarylether (L = O) to a 3-*H*-imidazo[1,2-*a*]pyridine was even used to efficiently prepare biarylethers **7** (L = O). Compounds **7z** and **7aa** were made by alkylation with benzyl bromides **15**. Finally, two biarylsulfones were prepared by coupling of 4-bromo-1-iodobenzene to imidazo[1,2-*a*]pyridine **10c** (Y = 2-Pr, Z = CF<sub>3</sub>) to give **18** followed by Suzuki coupling with methylsulfonylbenzeneboronic acids to provide **19a** and **19b** (Scheme 5).

To determine the affinity of imidazo[1,2-a]pyridines for the two LXR subtypes, binding assays were run using recombinant human ligand binding domains (LBDs) of the respective LXR $\alpha$  and LXR $\beta$  subtypes, measuring displacement of [ ${}^{3}$ H]T0901317 from the LBD (Table 1). ${}^{25}$  Comparing the simpler analogs **5a**, **6a**, and **7a** from the quinoline, benzimidazole, and imidazo[1,2-a]pyridine series, respectively, there was a significant loss in affinity (almost 50-fold vs **5a** for LXR $\beta$ ). The affinity did not improve with larger sulfones as the analogs **7b**–**7d** had similar affinity relative to **7a**. However, extensive exploration of the 2-substituent (Y) indicated it played an important role in LXR affinity in the series. While slightly larger groups like ethyl (**7e**), isopropyl (**7f**), and t-butyl (**7g**) and even larger phenyl (**7h**) had comparable affinity for LXR $\beta$ , enlarging to a benzyl group (**7i**) provided essentially the same affinity as seen with **5a**. While introduction of dimethylamine at the 2-position

**Scheme 1.** Reagents and conditions: (a) EtOH, reflux, 16–24 h; (b) Pd(OAc)<sub>2</sub> (0.02 equiv), Ph<sub>3</sub>P (0.04 equiv), Cs<sub>2</sub>CO<sub>3</sub> (1.0 equiv), dioxane, reflux, 16–24 h; (c) 20% Pd(OH)<sub>2</sub> on carbon (0.1 equiv), KOAc (3.0 equiv), DMA, 145 °C, 16–24 h; d) **12** (Hal = Br, I,) (1.0–2.0 equiv), Cul (0.1 equiv), Me<sub>2</sub>NCH<sub>2</sub>CO<sub>2</sub>H hydrochloride (0.375 equiv), Cs<sub>2</sub>CO<sub>3</sub> (3.0 equiv), dioxane, 100–110 °C, 16–24 h; (e) **12** (Hal = F, Cl) (1.0–2.0 equiv), K<sub>2</sub>CO<sub>3</sub> (1.0–2.0 equiv), DMF, 120–150 °C, 16–24 h; (f) TFA, CH<sub>2</sub>Cl<sub>2</sub>, 20 °C, 18–24 h.

**Scheme 2.** Reagents and conditions: (a)  $PhB(OH)_2$  (2.0 equiv),  $K_2CO_3$  (3.0 equiv),  $Pd(PPh_3)_4$  (0.03 equiv), dioxane, water, reflux, 1.5 h (51%).

**Scheme 3.** Reagents and conditions: (a) *N*-bromosuccinimide (1.1 equiv),  $\alpha,\alpha$ -azobisisobutyronitrile (0.1 equiv), MeCN, reflux, 3 h (79%); (b) azacycle (3.0–5.0 equiv), DMSO or ethanol, rt, 1–24 h (37–68%).

**Scheme 4.** Reagents and conditions: (a) 3-(MeSO<sub>2</sub>)PhB(OH)<sub>2</sub> (1 equiv), Pd(PPh<sub>3</sub>)<sub>4</sub> (0.03 equiv), 1 M aqueous Na<sub>2</sub>CO<sub>3</sub> (0.12 equiv), EtOH, PhMe, 80 °C, 18–24 h (8%); (b) **11a** (Y = 2-Pr, Z = CF<sub>3</sub>, 0.83 equiv), Cs<sub>2</sub>CO<sub>3</sub> (1.16 equiv), DMF, 40 °C, 20 h (96–99%); (c) 3-HO-PhSO<sub>2</sub>Me, K<sub>2</sub>CO<sub>3</sub>, DMF, 80 °C, 24 h (82%); (d) **10b** (Y = Et, Z = CF<sub>3</sub>), 20% Pd(OH)<sub>2</sub> on carbon (0.1 equiv), KOAc (3.0 equiv), DMA, 145 °C, 24 h (67–87%).

via a methylene linker gave much weaker affinity (7k), larger, more lipophilic azacycles (7l, 7m) trended toward better binding. The much more weakly basic amine found in thiazolidine gave a compound with excellent affinity (7n, hLXR $\beta$  IC $_{50}$  = 2.0 nM). However, imidazole in place of phenyl lost 30-fold in affinity (7o). Returning to a 2-isopropyl group and varying the Z-substituent, affinity decreased slightly in a chloro for trifluoromethyl exchange (7f vs 7p) while cyano fared much worse (7q). The importance of the hydrogen bond accepting ability of the sulfone can be inferred from the dramatic reduction in affinity (over 50-fold) seen with the trifluoromethylsulfone 7r compared to methylsulfone 7f, suggesting the importance of more electron rich sulfone oxygens. The receptors were more tolerant of large lipophilic groups over primary sulfonamides as seen in a comparison of 7t-7u. Interestingly, a secondary sulfonamide could also substitute for a sulfone

**Scheme 5.** Reagents and conditions: (a) 4-BrPhI, Pd(OAc) $_2$  (0.02 equiv), Ph $_3$ P (0.04 equiv), Cs $_2$ CO $_3$  (1.0 equiv), dioxane, reflux, 16–24 h; (b) MeSO $_2$ PhB(OH) $_2$  (1.3 equiv), Pd(PPh $_3$ ) $_4$  (0.15 equiv), Na $_2$ CO $_3$  (2.3 equiv), aqueous diglyme, 80 °C, 18–24 h.

as shown by **7v** which had similar binding affinity to **7p**, its closest analog. A methylene linker in place of oxygen lost sixfold in affinity (**7e** vs **7x**) while other linkers provided even weaker LXR ligands (**7y–7aa**). *para*-Biarylsulfones **19a** and **19b** had moderate and very weak LXR affinity, respectively.

Binding selectivity for LXR $\beta$  over LXR $\alpha$  subtypes was only moderate, though **7e**, **7h**, and **7p** showed selectivity (LXR $\alpha$  IC<sub>50</sub>/LXR $\beta$  IC<sub>50</sub>) of greater than tenfold. However, the in vivo functional implications of this level of binding selectivity are unclear. Essentially none of compounds had PPAR agonism when tested in PPAR functional assays reported earlier.<sup>12a</sup>

LXR functional activity was first examined in Gal4 assays, 10 transient transactivation assays in Huh7 cells transfected with human LXR ligand binding domains (LBDs) fused to Gal4 DNA binding domains. The assays were performed for both LXR $\alpha$  and LXR $\beta$  LBDs. As in the binding affinity, the potency in the Gal4 assays for 7a was weaker compared to the analogous benzimidazole 6a and much weaker compared to quinoline **5a**. Efficacy was also reduced, especially compared to **7a**. Larger sulfone groups gave comparable or weaker potency (7b-7d) while more lipophilic Y groups had little impact until a benzyl group was used (7i). Compound 7i was the most potent analog in this assay (Gal4 $\beta$  EC<sub>50</sub> = 20 nM) and was essentially as efficacious as 1. Saturated azacycles 7m and particularly 7n demonstrated good activity in the assay, with 7n being nearly as potent and efficacious as 7i. Both were more potent than 1. Sulfonamides 7t and 7v were good agonists in the assay while the methylene (7x) and oxymethylene (7z) spacers were much weaker compared to the biarylether analogs. Rather modest functional selectivity for LXRB over LXRa was seen for a few compounds particularly 7i and 7m, about sevenfold and fivefold selective, respectively. As noted above, none of the biarylethers tested had any PPAR activity.

Additional cell based assays tested for upregulation of ABCA1 mRNA in a THP1 human macrophage cell line  $^{25}$  and for an increase in cellular triglyceride levels in a HepG2 (human liver) cell line (Table 2), with increases in ABCA1 mRNA and in TG levels using 1 taken as 100% efficacy. Ideally, good candidates for antiatherosclerotic agents would show an ABCA1 mRNA increase with no increase in TG accumulation. Most of imidazo[1,2-a]pyridines 7 were about 10-fold weaker at increasing ABCA1 levels compared to 5a, with the exception of 7n which was essentially equipotent. However, little selectivity for ABCA1 mRNA increases relative to TG accumulation was observed. About a fourfold selectivity in potency was found for 7n, but this was less than seen for 5a. Higher concentrations (3  $\mu$ M and 10  $\mu$ M) of both 7a and 7b led to decreases in TG levels, an effect possibly related to cell toxicity affecting TG synthesis.

In an assay testing for stability in mouse, rat and human microsomes, compounds with smaller sulfones and 2-substituents (7a, 7b, 7e) had good stability while larger 2-substituents

Table 1 Biarylether sulfone imidazo[1,2-a]pyridines 7, 19<sup>a</sup>

	L	Х	Y	Z	LXRβ IC <sub>50</sub> (nM)	LXRα IC <sub>50</sub> (nM)	Gal4β EC <sub>50</sub> (μM) (% ag)	Gal4α EC <sub>50</sub> (μM) (% ag)
1		_	_	_	9	13	0.170 (100%)	0.135 (100%)
5a	_	m-SO <sub>2</sub> Me	Me	CF <sub>3</sub>	1.0	2.4	0.031 (78%)	0.183 (75%)
6a	_	m-SO <sub>2</sub> Me	Me	CF <sub>3</sub>	7.0	128	0.162 (49%)	0.64 (42%)
7a	0	m-SO <sub>2</sub> Me	Me	CF <sub>3</sub>	49	799	0.84 (37%)	2.54 (25%)
7b	0	m-SO <sub>2</sub> Et	Me	CF <sub>3</sub>	30	561	0.68 (39%)	2.28 (26%)
7c	0	m-SO <sub>2</sub> (i-Pr)	Me	$CF_3$	33	960	0.70 (25%)	4.1 (17%)
7d	0	m-SO <sub>2</sub> (CH <sub>2</sub> ) <sub>3</sub> OH	Me	$CF_3$	188	>1000	1.42 (17%)	6.0 (8%)
7e	0	m-SO <sub>2</sub> Me	Et	$CF_3$	14	217	0.42 (62%)	1.22 (58%)
7f	0	m-SO <sub>2</sub> Me	i-Pr	$CF_3$	3.5	137	0.95 (68%)	1.86 (44%)
7g	0	m-SO <sub>2</sub> Me	t-Bu	$CF_3$	6.5	222	0.71 (71%)	2.18 (55%)
7h	0	m-SO <sub>2</sub> Me	Ph	$CF_3$	7.2	91	0.57 (64%)	1.11 (54%)
7i	0	m-SO <sub>2</sub> Me	CH <sub>2</sub> Ph	$CF_3$	0.81	5.9	0.020 (88%)	0.144 (62%)
7j	0	m-SO <sub>2</sub> Me	CH <sub>2</sub> Br	$CF_3$	nt	nt	nt	nt
7k	0	m-SO <sub>2</sub> Me	$CH_2NMe_2$	$CF_3$	>1000	>1000	nt	nt
71	0	m-SO <sub>2</sub> Me	CH <sub>2</sub> -1-pyrrolidine	$CF_3$	279	>1000	nt	nt
7m	0	m-SO <sub>2</sub> Me	CH <sub>2</sub> -1-piperidine	$CF_3$	69	>1000	0.204 (56%)	0.92 (45%)
7n	0	m-SO <sub>2</sub> Me	CH <sub>2</sub> -3-thiazolidine	$CF_3$	2.0	9	0.032 (62%)	0.170 (64%)
<b>7o</b>	0	m-SO <sub>2</sub> Me	CH <sub>2</sub> -1-imidazole	CF <sub>3</sub>	30	365	0.85 (81%)	1.7 (79%)
7p	0	m-SO <sub>2</sub> Me	i-Pr	Cl	10	304	0.54 (50%)	1.66 (53%)
7q	0	m-SO <sub>2</sub> Me	i-Pr	CN	43	>1000	1.68 (44%)	3.9 (20%)
7r	0	m-SO <sub>2</sub> CF <sub>3</sub>	i-Pr	$CF_3$	194	>1000	nt	nt
7s	0	m-SO <sub>2</sub> N(PMB) <sub>2</sub>	i-Pr	Cl	nt	nt	nt	nt
7t	0	m-SO <sub>2</sub> NMe(PMB)	i-Pr	Cl	24	71	0.89 (48%)	0.93 (47%)
7u	0	m-SO <sub>2</sub> NH <sub>2</sub>	i-Pr	Cl	136	>1000	nt	nt
7v	0	m-SO <sub>2</sub> NHMe	i-Pr	Cl	7.5	239	0.53 (61%)	1.61 (46%)
7w	0	m-CH <sub>2</sub> SO <sub>2</sub> Me	i-Pr	$CF_3$	187	>1000	nt	nt
7x	$CH_2$	m-SO <sub>2</sub> Me	Et	$CF_3$	79	>1000	1.57 (48%)	3.2 (30%)
7у	$CH_2O$	m-SO <sub>2</sub> Me	Et	$CF_3$	292	>1000	nt	nt
7z	$OCH_2$	m-SO <sub>2</sub> Me	i-Pr	$CF_3$	89	>1000	3.24 (26%)	6.5 (9%)
7aa	$OCH_2$	p-SO <sub>2</sub> Me	i-Pr	$CF_3$	>1000	>1000	nt	nt
19ª	-	m-SO <sub>2</sub> Me	i-Pr	$CF_3$	166	>1000	na	na
19b	_	p-SO <sub>2</sub> Me	i-Pr	CF <sub>3</sub>	>1000	>1000	na	na

<sup>&</sup>lt;sup>a</sup> Results are given as the mean of two independent experiments. The standard deviations for binding assays were typically ±30% of mean or less. The standard deviations for the Gal4 assays were typically ±30% of the mean or less. % of efficacy is relative to 1. nt = not tested na = inactive. PMB = 4-MeOPhCH<sub>2</sub>.

Table 2 Cell-based assays, solubility and microsomal stability

	R	Y	ABCA1 THP1 EC <sub>50</sub> <sup>a</sup> (μM) (% eff)	Hepatocyte TG accum assay EC <sub>50</sub> <sup>a</sup> (μM) (% eff)	Solubility (μg/mL)	Microsomal stability, $t_{1/2}$ m, r, h, (min)	C <sub>max</sub> <sup>c</sup> (ng/mL)/AUC <sub>0-</sub> last (ng h/mL)	t <sub>max</sub> <sup>c</sup> (h)/ t <sub>1/2</sub> (h)
1	_	_	0.044 (100%)	0.137 (100%)	_	30, 30, 30	_	_
5a	Me	Me	0.010 (82%)	0.147 (69%)	<1	30, 30, 30	457/1690	2/nd
6a	Me	Me	0.42 (126%)	0.270 (37%)	214	30, 30, 30	614/8938	2/13.5
7a	Me	Me	0.125 (99%)	0.072 (8%) <sup>b</sup>	>100	30, 30, 30	1136/4667	0.25/4.5
7b	Et	Me	0.130 (100%)	nd <sup>b</sup>	68	30, 30, 21	1421/7792	1.0/6.1
7e	Me	Et	0.243 (65%)	0.227 (38%)	228	20, 30, 30	- '	<u> </u>
7i	Me	CH <sub>2</sub> Ph	0.12 (105%)	0.065 (81%)	3	2, 7, 19	_	_
7m	Me	CH <sub>2</sub> -1- piperidine	0.044 (100%)	0.128 (26%)	73	2, 5, 3	_	_
7n	Me	CH <sub>2</sub> -3- thiazolidine	0.010 (98%)	0.042 (36%)	1	3, 23, 6	_	_

<sup>&</sup>lt;sup>a</sup> Results are given as the mean of at least two independent experiments. The EC<sub>50</sub> values for the assays were typically ±50% of mean or less (variability for 7i was higher). % efficacy is relative to **1**.

<sup>b</sup> TG levels decreased at higher concentrations. nd = not determined.

<sup>&</sup>lt;sup>c</sup> C57 mouse PK dosing: 10 mg/kg po (gavage) in 0.5% methylcellulose/2% Tween in water, except **5a** dosed at 2.5 mg/kg po (gavage).

(**7i**, **7m**, **7n**) significantly reduced the half-life (Table 2). Solubility followed the same trends, increasing for smaller, less lipophilic compounds. In mice dosed orally (gavage) with several compounds, two imidazo[1,2-a]pyridines, **7a** and **7b**, gave comparable or slightly better  $C_{\text{max}}$  and exposure compared to representative examples from earlier series.

Compounds 7 in which an imidazo[1,2-a]pyridine replaced a quinoline or benzimidazole found in earlier biarylether sulfones led to a new series of LXR agonists. Efficient routes to synthesize 7 were developed, highlighted by a direct arylation reaction of imidazo[1,2-a]pyridines 10 with an arylbromide or aryliodide. A lipophilic 2-substituent was important for both good affinity and functional activity in Gal4 and ABCA1 mRNA gene expression assays. Little selectivity for desired increases in ABCA1 lipid transporter relative to undesired TG accumulation was found.

### Acknowledgments

We thank the Wyeth Discovery Analytical Chemistry Department for analytical data and Anita Halpern and Dawn Savio for biological assay data. We acknowledge the contributions and support of Drs. Li Di, Jeremy Travins, Ronald Magolda (deceased June 1, 2008), Tarek Mansour, Steven Gardell, and George Vlasuk.

### **References and notes**

- (a) Fievet, C.; Staels, B. Biochem. Pharmacol. 2009, 77, 1316; (b) Goodwin, B. J.;
   Zuercher, W. J.; Collins, J. L. Curr. Top. Med. Chem. 2008, 8, 781.
- 2. For a recent review: Baldan, A.; Bojanic, D.; Edwards, P. A. J. Lipid Res. 2009, 50, S80
- LXR agonists also activate the carbohydrate-response element-binding protein (ChREBP) which may contribute to steatosis: Cha, J.-Y.; Repa, J. J. J. Biol. Chem. 2006. 282, 743.
- 4. Quinet, E.; Savio, D. A.; Halpern, A. R.; Chen, L.; Schuster, G. U.; Gustafsson, J.-A.; Basso, M. D.; Nambi, P. Mol. Pharmacol. 2006, 70, 1340. and Ref. 1a.
- Repa, J. J.; Turley, S. D.; Lobaccaro, J. A.; Medina, J.; Li, L.; Lustig, K.; Shan, B.; Heyman, R. A.; Dietschy, J. M.; Mangelsdorf, D. J. Science 2000, 289, 1524; Li, L. et al Bioorg. Med. Chem. Lett. 2006, 16, 1638.
- Collins, J. L. et al J. Med. Chem. 2002, 45, 1963; See also: Marino, J. P.; Kallander, L. S.; Ma, C.; Oh, H.-J.; Lee, D.; Gaitanopoulos, D. E.; Krawiec, J. A.; Parks, D. J.; Webb, C. L.; Ziegler, K.; Jaye, M.; Thompson, S. K. Bioorg. Med. Chem. Lett. 2009, 19, 5617.
- 7. Wrobel, J.; Steffan, R.; Bowen, S. M.; Magolda, R.; Matelan, E.; Unwalla, R.; Basso, M.; Clerin, V.; Gardell, S. J.; Nambi, P.; Quinet, E.; Reminick, J. I.; Vlasuk, G. P.; Wang, S.; Feingold, I.; Huselton, C.; Bonn, T.; Farnegardh, M.; Hansson, T.;

- Goos-Nilsson, A.; Wilhelmsson, A.; Zamaratski, E.; Evans, M. J. *J. Med. Chem.* **2008**, *51*, 7161.
- Quinet, E.; Basso, M. D.; Halpern, A. R.; Steffan, R. J.; Yates, D. W.; DiBlasio-Smith, E.; Mounts, W. M.; LaVallie, E.; Wrobel, J.; Nambi, P. Abstract 585: A Novel LXR Ligand Reduces LDL Cholesterol in Monkey Circulation, 2007, 116: II, 106
- 9. Katz, A.; Udata, C.; Ott, E.; Hickey, L.; Burczynski, M. E.; Burghart, P.; Vesterqvist, O.; Meng, X. J. Clin. Pharmacol. 2009, 49, 643.
- Hu, B.; Collini, M.; Unwalla, R.; Miller, C.; Singhaus, R.; Quinet, E.; Savio, D.; Halpern, A.; Basso, M.; Keith, J.; Clerin, V.; Chen, L.; Resmini, C.; Liu, Q.-Y.; Feingold, I.; Huselton, C.; Azam, F.; Farnegardh, M.; Enroth, C.; Bonn, T.; Goos-Nilsson, A.; Wilhelmsson, A.; Nambi, P.; Wrobel, J. J. Med. Chem. 2006, 49, 6151.
- Hu, B.; Quinet, E.; Unwalla, R.; Collini, M.; Jetter, J.; Dooley, R.; Andraka, D.; Nogle, L.; Savio, D.; Halpern, A.; Goos-Nilsson, A.; Wilhelmsson, A.; Nambi, P.; Wrobel, J. Bioorg. Med. Chem. Lett. 2008, 18, 54. and Ref. 12.
- (a) Bernotas, R.; Singhaus, R.; Kaufman, D.; Ullrich, J.; Fletcher, H.; Quinet, E.; Nambi, P.; Unwalla, R.; Savio, D.; Wilhelmsson, A.; Goos-Nilsson, A.; Farnegardh, M.; Wrobel, J. Bioorg. Med. Chem. 2009, 17, 1663; (b) Bernotas, R.; Kaufman, D.; Singhaus, R.; Ullrich, J.; Unwalla, R.; Quinet, E.; Nambi, P.; Wilhelmsson, A.; Goos-Nilsson, A.; Wrobel, J. Bioorg. Med. Chem. 2009, 17, 8086.
- Bernotas, R. C.; Singhaus, R. R.; Kaufman, D. H.; Travins, J. T.; Ullrich, J. W.; Unwalla, R. J.; Quinet, E.; Evans, M.; Nambi, P.; Olland, A.; Kauppi, B.; Wilhelmsson, A.; Goos-Nilsson, A.; Wrobel, J. Bioorg. Med. Chem. Lett., in press. doi:10.1016/j.bmcl.2009.10.132.
- Bernotas, R. C.; Travins, J. M.; Wrobel, J. E.; Kaufman, D. H. Patent WO 2009086138 A1 and Travins, J. M.; Bernotas, R. C.; Kaufman, D. H.; Quinet, E.; Nambi, P.; Feingold, I.; Huselton, C.; Wilhelmsson, A.; Goos-Nilsson, A.; Wrobel, J. Bioorg. Med. Chem. Lett., in press. doi:10.1016/j.bmcl.2009.11.099.
- New compounds gave satisfactory <sup>1</sup>H NMR, MS, and HRMS data. Compounds were generally >95% pure by HPLC analysis. Further experimental details are given in patent application WO 2009086123.
- 6. Barlin, G. B.; Davies, L. P.; Harrison, P. W. Aust. J. Chem. 1995, 48, 1031.
- 17. Chowdhury, S.; Georghiou, P. E. Tetrahedron Lett. 1999, 40, 7599.
- Koubachi, J.; El Kazzouli, S.; Berteina-Raboin, S.; Mouaddib, A.; Guillaumet, G. Synlett 2006, 3237.
- 19. Parisien, M.; Valette, D.; Fagnou, K. J. Org. Chem. 2005, 70, 7578.
- For sulfones: (a) Zhu, W.; Ma, D. J. Org. Chem. 2005, 70, 2696; (b) Webb, K. Tetrahedron Lett. 1994, 35, 3457; (c) Antane, S.; Bernotas, R.; Li, Y.; McDevitt, R.; Yan, Y. Synth. Commun. 2004, 34, 2443; (d) Ragni, R.; Orselli, E.; Kottas, G. S.; Omar, O. H.; Babudri, F.; Pedone, A.; Naso, F.; Farinola, G. M.; De Cola, L. Chem. Eur. J. 2009, 15, 136. For sulfonamides, see patent application WO20090866123.
- 21. Ma, D.; Cai, Q. *Org. Lett.* **2003**, *5*, 3799; For alternative biarylether synthesis procedures, see: Evans, D. A.; Katz, J. L.; West, T. R. *Tetrahedron Lett.* **1998**, 39, 2937; Chan, D. M. T.; Monaco, K. L.; Wang, R.-P.; Winters, M. P. *Tetrahedron Lett.* **1998**, *39*, 2933.
- Fotsch, C.; Sonnenberg, J. D.; Chen, N.; Hale, C.; Karbon, W.; Norman, M. H. J. Med. Chem. 2001, 44, 2344.
- For an example of para-methoxybenzyl groups for the protection of sulfonamides, see: Evans, P. A.; Robinson, J. E. Org. Lett. 1999, 1, 1929.
- 24. Langle, S.; Abarbri, M.; Duchene, A. Tetrahedron Lett. 2003, 44, 9255.
- Quinet, E. M.; Savio, D. A.; Halpern, A. R.; Chen, L.; Miller, C. P.; Nambi, P. J. Lipid Res. 2004, 45, 1929.